# **Original Article**

Open camera or QR reader and scan code to access this article and other resources online.



# Epitope Mapping of an Anti-CD44v4 Monoclonal Antibody (C<sub>44</sub>Mab-108) Using Enzyme-Linked Immunosorbent Assay

Hiroyuki Suzuki, Mayuki Tawara, Aoi Hirayama, Nohara Goto, Tomohiro Tanaka, Mika K. Kaneko, and Yukinari Kato

CD44 is a type I transmembrane glycoprotein and possesses various isoforms which are largely classified into CD44 standard (CD44s) and CD44 variant (CD44v) isoforms. Some variant-encoded regions play critical roles in tumor progression. However, the function of CD44 variant 4 (CD44v4)-encoded region has not been fully understood. Using peptide immunization, we developed an anti-CD44v4 monoclonal antibody,  $C_{44}$ Mab-108, which is useful for flow cytometry, western blotting, and immunohistochemistry. In this study, we determined the critical epitope of  $C_{44}$ Mab-108 by enzyme-linked immunosorbent assay (ELISA). We used the alanine (or glycine)-substituted peptides of the CD44v4-encoded region (amino acids 271–290 of human CD44v3-10) and found that  $C_{44}$ Mab-108 did not recognize the alanine-substituted peptides of D280A and W281A. Furthermore, these peptides could not inhibit the recognition of  $C_{44}$ Mab-108 in flow cytometry and immunohistochemistry. The results indicate that the critical binding epitope of  $C_{44}$ Mab-108 includes Asp280 and Trp281 of CD44v3-10.

Keywords: CD44, CD44 variant 4, monoclonal antibody, epitope, enzyme-linked immunosorbent assay

### Introduction

C D44 HAS VARIOUS isoforms, which are generated by the alternative splicing of CD44 pre-mRNA.<sup>1</sup> The mRNA of CD44 standard (CD44s) isoform is produced by constant region exons, including the first five (1–5) and the last five (16–20).<sup>2</sup> The mRNAs of CD44 variant (CD44v) isoform are produced by the assembling of variant exons (v1–v10) with the constant region exons of CD44s.<sup>3</sup> CD44s and CD44v receive the posttranslational modifications, including *N*- or *O*-glycosylation.<sup>4</sup> Both CD44s and CD44v can attach to hyaluronic acid, which is important for cellular adhesion, homing, and motility.<sup>5</sup>

CD44v plays important roles in the tumor progression by specific functions of variant exon-encoded regions.<sup>6</sup> The heparin-binding growth factors are recruited to heparan sulphate modified in the v3-encoded region.<sup>7,8</sup> MET, a receptor tyrosine kinase for hepatocyte growth factor, associates with the v6-encoded region.<sup>9,10</sup> These functions are essential for the activation of growth factor signaling and tumor proliferation. However, the roles of CD44 variant 4 (CD44v4)-encoded

region have not been investigated. Therefore, specific antibodies against CD44v4 are indispensable for basic research, tumor diagnosis, and therapy.

We have established anti-CD44 monoclonal antibodies (mAbs), which recognize the standard<sup>11,12</sup>—or each variant<sup>13–20</sup>—encoded region. All mAbs can be used for flow cytometry, western blotting, and immunohistochemistry and are expected to contribute not only to basic research but also to phathological diagnosis. We also determined the critical epitopes of C<sub>44</sub>Mab-5<sup>21</sup> and C<sub>44</sub>Mab-46.<sup>22,23</sup> Clinical trials of anti-CD44 mAbs have been con-

Clinical trials of anti-CD44 mAbs have been conducted.<sup>24</sup> A humanized anti-CD44v6 mAb BIWA4 (bivatuzumab)–mertansine drug conjugate was evaluated, but discontinued due to the severe skin toxicities.<sup>25,26</sup> An antipan CD44 mAb, RG7356, exhibited an acceptable safety profile in patients with advanced CD44-expressing solid tumors. However, the study was terminated due to no evidence of a clinical and pharmacodynamic dose–response relationship with RG7356.<sup>27</sup> Therefore, the development of anti-CD44 mAbs with more potent and fewer side effects is required.

Department of Antibody Drug Development, Tohoku University Graduate School of Medicine, Sendai, Japan. This paper was previously published in preprint.org (doi: 10.20944/preprints202311.0216.v1).

We previously established an anti-CD44v4 mAb,  $C_{44}$ Mab-108 (IgG<sub>1</sub>, kappa) using the peptide immunization.<sup>19</sup> To clarify further characteristics of  $C_{44}$ Mab-108, we performed epitope mapping using enzyme-linked immunosorbent assay (ELISA).

#### **Materials and Methods**

## Peptides

The CD44v4 peptide (<sub>271</sub>-AFDHTKQNQDWTQWNPS HSN-<sub>290</sub>) and 20 alanine (or glycine)-substituted peptides (Table 1) were synthesized by utilizing PEPscreen (Sigma-Aldrich Corp., St. Louis, MO, USA). The number of amino acids (aa) is derived from human CD44v3-10 (Accession No.: X66733).

# ELISA

The CD44v4 peptides were immobilized on Nunc Maxisorp 96-well immunoplates (Thermo Fisher Scientific, Inc., Waltham, MA, USA) at a concentration of 10  $\mu$ g/mL for 30 minutes at 37°C. After washing with phosphate-buffered saline (PBS) containing 0.05% Tween20 (PBST; Nacalai Tesque, Inc., Kyoto, Japan), wells were blocked with 1% bovine serum albumin (BSA)-containing PBST for 30 minutes at 37°C. The plates were incubated with 10  $\mu$ g/mL of C<sub>44</sub>Mab-108, followed by a peroxidase-conjugated antimouse immunoglobulins (1:2000 diluted; Agilent Technologies, Inc., Santa Clara, CA, USA). Enzymatic reactions were performed using the ELISA POD Substrate TMB Kit (Nacalai Tesque, Inc.). Optical density was measured at 655 nm using an iMark microplate reader (Bio-Rad Laboratories, Inc., Berkeley, CA, USA).

#### TABLE 1. IDENTIFICATION OF THE C<sub>44</sub>MAB-108 EPITOPE USING ALANINE (OR GLYCINE)-SUBSTITUTED CD44 VARIANT 4 PEPTIDES

Peptides	Sequences	C44Mab-108
WT (271, 290)	AFDHTKQNQDWTQWNPSHSN	+++
A271G	GFDHTKONODWTOWNPSHSN	+++
F272A	AADHTKÒNÒDWTÒWNPSHSN	+++
D273A	AFAHTKONODWTOWNPSHSN	+++
H274A	AFDATKÕNÕDWTÕWNPSHSN	+++
T275A	AFDHAKÒNÒDWTÒWNPSHSN	+++
K276A	AFDHTAONODWTOWNPSHSN	+++
O277A	AFDHTKÀNÒDWTÒWNPSHSN	+++
N278A	AFDHTKOAODWTOWNPSHSN	+++
O279A	AFDHTKÕNÄDWTÕWNPSHSN	+++
D280A	AFDHTKÕNOAWTÕWNPSHSN	_
W281A	AFDHTKÕNÕDATÕWNPSHSN	_
T282A	AFDHTKÒNÒDWAÒWNPSHSN	+
Q283A	AFDHTKQNQDWTAWNPSHSN	+++
Ŵ284A	AFDHTKQNQDWTQANPSHSN	+
N285A	AFDHTKÕNÕDWTÕWAPSHSN	+++
P286A	AFDHTKÕNÕDWTÕWNASHSN	+++
S287A	AFDHTKÕNÕDWTÕWNPAHSN	+++
H288A	AFDHTKÕNÕDWTÕWNPSASN	+++
S289A	AFDHTKONODWTOWNPSHAN	+++
N290A	AFDHTKQNQDWTQWNPSHSA	+++

+++, OD655 $\geq$ 0.6; ++, 0.3  $\leq$  OD655<0.6; +, 0.1  $\leq$  OD655<0.3; -, OD655<0.1.

#### Flow cytometry

The N-terminal PA16-tagged CD44v3-10-overexpressed Chinese hamster ovary-K1 (CHO/CD44v3-10)<sup>19</sup> cells were harvested after a brief exposure to 0.25% trypsin in 1 mM ethylenediaminetetraacetic acid (EDTA; Nacalai Tesque, Inc.) and washed with 0.1% BSA in PBS. C<sub>44</sub>Mab-108 (10  $\mu$ g/mL) was incubated with the CD44v4 peptides (10  $\mu$ g/mL) for 30 minutes at 4°C. The cells were further treated with Alexa Fluor 488-conjugated anti-mouse IgG (1:2000). Fluorescence data were collected using the SA3800 Cell Analyzer (Sony Corp., Tokyo, Japan).

#### Immunohistochemical analysis

One formalin-fixed paraffin-embedded (FFPE) tissue of oral squamous cell carcinoma (OSCC) for peptide blocking assay was obtained from Tokyo Medical and Dental University.<sup>28</sup> The tissue slides were autoclaved in citrate buffer (pH 6.0; Nichirei Biosciences, Inc., Tokyo, Japan) for 20 minutes for antigen retrieval. After blocking with Super Block T20 (Thermo Fisher Scientific, Inc.), the sections were incubated with C<sub>44</sub>Mab-108 (10  $\mu$ g/mL) in the presence or absence of the CD44v4 peptides (10  $\mu$ g/mL) and then treated with the EnVision+ Kit for mouse (Agilent Technologies, Inc.) for 30 minutes. The color was developed using 3,3'-diaminobenzidine tetrahydrochloride (DAB; Agilent Technologies, Inc.). Counterstaining was performed with hematoxylin (FUJIFILM Wako Pure Chemical Corporation, Osaka, Japan). Leica DMD108 (Leica Microsystems GmbH, Wetzlar, Germany) was used to examine the sections and obtain images.

## Results

## Epitope mapping of C<sub>44</sub>Mab-108 with alanine (or glycine)-substituted CD44v4 peptides

We previously established an anti-CD44v4 mAb (C44Mab-108) by peptide immunization of CD44v4 region (273-DHTKQNQDWTQWNPSHSNP-291).<sup>19</sup> We confirmed that C44Mab-108 recognizes only the variant 4-encoded region peptide (aa 271-290), but not other regions of CD44v3-10 extracellular domain.<sup>19</sup> To identify the binding epitope of C<sub>44</sub>Mab-108, we synthesized 20 alanine (or glycine)substituted peptides of the CD44v4 (Table 1). C<sub>44</sub>Mab-108 exhibited reaction with A271G, F272A, D273A, H274A, T275A, K276A, Q277A, N278A, Q279A, T282A, Q283A, W284A, N285A, P286A, S287A, H288A, S289A, N290A, and wild type (WT) (Fig. 1A). In contrast, C44Mab-108 did not react with D280A and W281A (Fig. 1A). This result indicated that Asp280 and Trp281 are included in the critical epitope of C44Mab-108. The results are summarized in Table 1. Figure 1B shows the schematic illustration of CD44s, CD44v3-10, and the critical aa (Asp280 and Trp281) recognized by C<sub>44</sub>Mab-108.

# Flow cytometry using $C_{44}$ Mab-108 with alanine-substituted CD44v4 peptides

We next performed a peptide-blocking assay using flow cytometry to confirm the importance of the  $C_{44}$ Mab-108 epitope. As shown in Figure 2,  $C_{44}$ Mab-108 reacted with the CHO/CD44v3-10 cells. This reaction was completely neutralized by the WT peptide. In contrast, D280A and W281A

WT, wild type.



**FIG. 1.** Determination of the C<sub>44</sub>Mab-108 epitope by ELISA using alanine (or glycine)-substituted CD44v4 peptides. (**A**) The alanine (or glycine)-substituted CD44v4 peptides in PBS or PBS alone were immobilized on immunoplates. The plates were incubated with C<sub>44</sub>Mab-108 (10 µg/mL), followed by peroxidase-conjugated anti-mouse immunoglobulins. Error bars represent means ± SDs. N.C. (PBS). (**B**) Schematic illustration of CD44s, CD44v3-10, and the C<sub>44</sub>Mab-108 epitope. The C<sub>44</sub>Mab-108 epitope includes Asp280 and Trp281 of CD44v3-10. The epitope is conserved in mouse and Chinese hamster (Cham), but not in Rat. CD44s, CD44 standard; CD44v4, CD44 variant 4; ELISA, enzyme-linked immunosorbent assay; N.C., negative control; PBS, phosphate-buffered saline; SD, standard deviation; WT, wild type.

peptides did not block the reaction of  $C_{44}$ Mab-108 with CHO/CD44v3-10. The result confirmed that Asp280 and Trp281 of CD44v3-10 are critical for detection by  $C_{44}$ Mab-108 using flow cytometry.

# Immunohistochemistry using C<sub>44</sub>Mab-108 with alanine-substituted CD44v4 peptides

We also performed a peptide-blocking assay using immunohistochemical analysis. As shown in Figure 3,  $C_{44}$ Mab-108 stained the FFPE section of OSCC, which was completely neutralized by the WT peptide. In contrast, D280A and W281A peptides did not neutralize the reaction. These results were corresponding to that of Figure 2.

# Discussion

In this study, the critical epitope of  $C_{44}$ Mab-108 was determined as Asp280 and Trp281 in the CD44v4 region. Since the reactivity of  $C_{44}$ Mab-108 to T282A and W284A was also reduced (Fig. 1A), Thr282 and Trp284 may contribute to the



**FIG. 2.** Flow cytometry using C<sub>44</sub>Mab-108 and CD44v4 peptides. C<sub>44</sub>Mab-108 (10  $\mu$ g/mL) plus WT, the alanine-substituted peptides (10  $\mu$ g/mL), or control (0.1% DMSO in blocking buffer, –peptide) were reacted with CHO/CD44v3-10 cells for 30 minutes at 4°C, followed by treatment with Alexa Fluor 488-conjugated anti-mouse IgG. The black line represents the control. DMSO, dimethyl sulfoxide; WT, wild type.



**FIG. 3.** Immunohistochemistry using C<sub>44</sub>Mab-108 and CD44v4 peptides. The FFPE sections of OSCC were incubated with C<sub>44</sub>Mab-108 ( $10 \mu g/mL$ ) plus WT, the alanine-substituted peptides ( $10 \mu g/mL$ ), or control (0.1% DMSO in blocking buffer, –peptide), followed by that with the Envision + Kit. Scale bar= $100 \mu m$ . FFPE, formalin-fixed paraffin-embedded; OSCC, oral squamous cell carcinoma.

recognition partially. Figure 1B shows the homology of the v4 region among human, mouse, Chinese hamster, and rat sequences. The Asp280 and Trp281 are conserved in human, mouse, Chinese hamster, but not in rat. Furthermore, Thr282 and Trp284 are also conserved in human and mouse. Although the result suggests that  $C_{44}$ Mab-108 might recognize both human and mouse CD44v4, it did not react with mouse CD44v4 using flow cytometry (data not shown).

Since  $C_{44}$ Mab-108 was established by the peptide immunization,  $C_{44}$ Mab-108 can recognize the definite peptide structure of the variant 4-encoded region. In contrast, CD44 is predicted to carry 146 *O*-glycosylation sites in the variant region. Among them, 41 of these sites have already been experimentally confirmed.<sup>29</sup> Thr282 is also a confirmed *O*-glycosylation site.<sup>30</sup> Further studies are required to reveal whether the *O*-glycan at Thr282 affects the recognition by  $C_{44}$ Mab-108.

In our previous study,  $C_{44}$ Mab-108 could detect CD44v3-10-overexpressed cells such as CHO/CD44v3-10, but not detect endogenous CD44v4 in several cancer cell lines in flow cytometry.<sup>19</sup> In contrast,  $C_{44}$ Mab-108 could detect endogenous and membranous CD44v4 in immunohistochemistry.<sup>19</sup> These results suggest that the variant 4-encoded region is folded into the inside of protein in living cells, but exposed by antigen retrieval in immunohistochemistry. Recently, we provided a potential strategy for developing cancer-specific antibodies that target locally misfolded cell surface receptors such as human epidermal growth factor receptor 2.<sup>31,32</sup> Further studies are needed to investigate whether  $C_{44}$ Mab-108 is involved in the recognition of specific CD44v4 type and/or specific condition of cells.

#### Authors' Contributions

H.S., M.T., A.H., N.G., and T.T. performed the experiments. M.K.K. and Y.K. designed the experiments. H.S. and M.K.K. analyzed the data. H.S. and Y.K. wrote the manuscript. All authors have read and agreed to the published version of the manuscript.

#### **Author Disclosure Statement**

No competing financial interests exist.

#### **Funding Information**

This research was supported in part by Japan Agency for Medical Research and Development (AMED) under Grant Nos.: JP23ama121008 (to Y.K.), JP23am0401013 (to Y.K.), 23bm1123027h0001 (to Y.K.), and JP23ck0106730 (to Y.K.), and by the Japan Society for the Promotion of Science (JSPS) Grants-in-Aid for Scientific Research (KA-KENHI) Grant Nos. 22K06995 (to H.S.), 21K07168 (to M.K.K.), and 22K07224 (to Y.K.).

#### References

- Ponta H, Sherman L, Herrlich PA. CD44: From adhesion molecules to signalling regulators. Nat Rev Mol Cell Biol 2003;4:33–45; doi: 10.1038/nrm1004
- Yan Y, Zuo X, Wei D. Concise review: Emerging role of CD44 in cancer stem cells: A promising biomarker and therapeutic target. Stem Cells Transl Med 2015;4:1033– 1043; doi: 10.5966/sctm.2015-0048.

- Chen C, Zhao S, Karnad A, et al. The biology and role of CD44 in cancer progression: Therapeutic implications. J Hematol Oncol 2018;11:64; doi: 10.1186/s13045-018-0605-5
- 4. Liao C, Wang Q, An J, et al. CD44 glycosylation as a therapeutic target in oncology. Front Oncol 2022;12: 883831; doi: 10.3389/fonc.2022.883831
- Mishra MN, Chandavarkar V, Sharma R, et al. Structure, function and role of CD44 in neoplasia. J Oral Maxillofac Pathol 2019;23:267–272; doi: 10.4103/jomfp.JOMFP\_ 246\_18
- Zöller M. CD44: Can a cancer-initiating cell profit from an abundantly expressed molecule? Nat Rev Cancer 2011;11: 254–267; doi: 10.1038/nrc3023
- Bennett KL, Jackson DG, Simon JC, et al. CD44 isoforms containing exon V3 are responsible for the presentation of heparin-binding growth factor. J Cell Biol 1995;128:687– 698; doi: 10.1083/jcb.128.4.687
- Jackson DG, Bell JI, Dickinson R, et al. Proteoglycan forms of the lymphocyte homing receptor CD44 are alternatively spliced variants containing the v3 exon. J Cell Biol 1995; 128:673–685; doi: 10.1083/jcb.128.4.673
- Matzke A, Sargsyan V, Holtmann B, et al. Haploinsufficiency of c-Met in cd44–/– mice identifies a collaboration of CD44 and c-Met in vivo. Mol Cell Biol 2007;27:8797–8806; doi: 10.1128/mcb.01355-07
- Orian-Rousseau V, Chen L, Sleeman JP, et al. CD44 is required for two consecutive steps in HGF/c-Met signaling. Genes Dev 2002;16:3074–3086; doi: 10.1101/gad .242602
- Yamada S, Itai S, Nakamura T, et al. Detection of high CD44 expression in oral cancers using the novel monoclonal antibody, C(44)Mab-5. Biochem Biophys Rep 2018; 14:64–68; doi: 10.1016/j.bbrep.2018.03.007
- Goto N, Suzuki H, Tanaka T, et al. Development of a novel anti-CD44 monoclonal antibody for multiple applications against esophageal squamous cell carcinomas. Int J Mol Sci 2022;23:5535; doi: 10.3390/ijms23105535
- Ejima R, Suzuki H, Tanaka T, et al. Development of a novel anti-CD44 variant 6 monoclonal antibody C(44)Mab-9 for multiple applications against colorectal carcinomas. Int J Mol Sci 2023;24:4007; doi: 10.3390/ijms24044007
- Ishikawa K, Suzuki H, Kaneko MK, et al. Establishment of a novel anti-CD44 variant 10 monoclonal antibody C(44)Mab-18 for immunohistochemical analysis against oral squamous cell carcinomas. Curr Issues Mol Biol 2023; 45:5248–5262; doi: 10.3390/cimb45070333
- Kudo Y, Suzuki H, Tanaka T, et al. Development of a novel anti-CD44 variant 5 monoclonal antibody C(44)Mab-3 for multiple applications against pancreatic carcinomas. Antibodies (Basel) 2023;12:31; doi: 10.3390/antib12020031
- Suzuki H, Goto N, Tanaka T, et al. Development of a novel anti-CD44 variant 8 monoclonal antibody C(44)Mab-94 against gastric carcinomas. Antibodies (Basel) 2023;12:45; doi: 10.3390/antib12030045
- Suzuki H, Kitamura K, Goto N, et al. A novel anti-CD44 variant 3 monoclonal antibody C(44)Mab-6 was established for multiple applications. Int J Mol Sci 2023;24:8411; doi: 10.3390/ijms24098411
- Suzuki H, Ozawa K, Tanaka T, et al. Development of a novel anti-CD44 Variant 7/8 monoclonal antibody, C(44)Mab-34, for multiple applications against oral carcinomas. Biomedicines 2023;11:1099; doi: 10.3390/ biomedicines11041099

- Suzuki H, Tanaka T, Goto N, et al. Development of a novel anti-CD44 variant 4 monoclonal antibody C(44)Mab-108 for immunohistochemistry. Curr Issues Mol Biol 2023;45: 1875–1888; doi: 10.3390/cimb45030121
- Tawara M, Suzuki H, Goto N, et al. A novel anti-CD44 variant 9 monoclonal antibody C(44)Mab-1 was developed for immunohistochemical analyses against colorectal cancers. Curr Issues Mol Biol 2023;45:3658–3673; doi: 10 .3390/cimb45040238
- Asano T, Kaneko MK, Kato Y. Development of a novel epitope mapping system: RIEDL Insertion for Epitope Mapping Method. Monoclon Antib Immunodiagn Immunother 2021;40:162–167; doi: 10.1089/mab.2021.0023
- 22. Asano T, Kaneko MK, Takei J, et al. Epitope mapping of the Anti-CD44 monoclonal antibody (C(44)Mab-46) using the REMAP Method. Monoclon Antib Immunodiagn Immunother 2021;40:156–161; doi: 10.1089/mab.2021.0012
- Takei J, Asano T, Suzuki H, et al. Epitope mapping of the anti-CD44 monoclonal antibody (C(44)Mab-46) using alanine-scanning mutagenesis and surface plasmon resonance. Monoclon Antib Immunodiagn Immunother 2021; 40:219–226; doi: 10.1089/mab.2021.0028
- Orian-Rousseau V, Ponta H. Perspectives of CD44 targeting therapies. Arch Toxicol 2015;89:3–14; doi: 10.1007/ s00204-014-1424-2
- 25. Riechelmann H, Sauter A, Golze W, et al. Phase I trial with the CD44v6-targeting immunoconjugate bivatuzumab mertansine in head and neck squamous cell carcinoma. Oral Oncol 2008;44:823–829; doi: 10.1016/j.oraloncology.2007 .10.009
- 26. Tijink BM, Buter J, de Bree R, et al. A phase I dose escalation study with anti-CD44v6 bivatuzumab mertansine in patients with incurable squamous cell carcinoma of the head and neck or esophagus. Clin Cancer Res 2006;12: 6064–6072; doi: 10.1158/1078-0432.CCR-06-0910
- Menke-van der Houven van Oordt CW, Gomez-Roca C, van Herpen C, et al. First-in-human phase I clinical trial of RG7356, an anti-CD44 humanized antibody, in patients

with advanced, CD44-expressing solid tumors. Oncotarget 2016;7:80046–80058; doi: 10.18632/oncotarget.11098

- Itai S, Ohishi T, Kaneko MK, et al. Anti-podocalyxin antibody exerts antitumor effects via antibody-dependent cellular cytotoxicity in mouse xenograft models of oral squamous cell carcinoma. Oncotarget 2018;9:22480– 22497; doi: 10.18632/oncotarget.25132
- 29. Steentoft C, Vakhrushev SY, Joshi HJ, et al. Precision mapping of the human O-GalNAc glycoproteome through SimpleCell technology. Embo j. 2013;32:1478–1488; doi: 10.1038/emboj.2013.79
- Mereiter S, Martins ÁM, Gomes C, et al. O-glycan truncation enhances cancer-related functions of CD44 in gastric cancer. FEBS Lett 2019;593:1675–1689; doi: 10.1002/ 1873-3468.13432
- Arimori T, Mihara E, Suzuki H, et al. Locally misfolded HER2 expressed on cancer cells is a promising target for development of cancer-specific antibodies. Structure 2024; doi: 10.1016/j.str.2024.02.007
- 32. Kaneko MK, Suzuki H, Ohishi T, et al. A cancer-specific monoclonal antibody against HER2 exerts antitumor activities in human breast cancer xenograft models. Int J Mol Sci 2024;25:1941; doi: 10.3390/ijms25031941

Address correspondence to: Yukinari Kato Department of Antibody Drug Development Tohoku University Graduate School of Medicine 2-1, Seiryo-machi, Aoba-ku Sendai Miyagi 980-8575 Japan

E-mail: yukinari.kato.e6@tohoku.ac.jp

Received: November 7, 2023 Accepted: January 17, 2024