

Immunohistochemical Detection of Sheep Podoplanin Using an Antibovine Podoplanin Monoclonal Antibody PMAb-44

Yukinari Kato,^{1,2} Shinji Yamada,¹ Shunsuke Itai,¹ Atsushi Kobayashi,³ Satoru Konnai,^{4,5} and Mika K. Kaneko¹

Podoplanin (PDPN) obtained from various animal species has been characterized using specific anti-PDPN monoclonal antibodies (mAbs), namely, PMAb-1, PMAb-2, PMAb-32, PMAb-38, PMAb-44, and PMAb-52 against mouse, rat, rabbit, dog, bovine, and cat PDPN, respectively. PDPN is expressed in type I alveolar cells in lungs, lymphatic endothelial cells, and kidney podocytes. In this study, we investigated possible cross-reactions between anti-PDPN mAbs and sheep PDPN. Type I alveolar cells from sheep lung were strongly detected by PMAb-44 using immunohistochemical analyses. These results indicate that PMAb-44 may be useful for the detection of sheep PDPN.

Keywords: sheep podoplanin, sheep PDPN, PMAb-44

Introduction

PODOPLANIN (PDPN) is a type I transmembrane sialoglycoprotein that induces platelet aggregation through the C-type lectin-like receptor-2 (CLEC-2).⁽¹⁾ It comprises three platelet aggregation-stimulating (PLAG) domains, namely, PLAG1–PLAG3 (EDxxVTPG sequence).⁽²⁾ Previously, we have shown that PLAG3 (platelet aggregation-stimulating domain 3) is the most important domain for platelet aggregation mediated by human PDPN.^(1–3) Recently, PLAG4 (EDxxT sequence), a PLAG-like domain, was reported to be a critical sequence for the PDPN–CLEC-2 interaction.⁽⁴⁾ The small cytoplasmic C-terminus (nine amino acids) interacts with members of the ERM (ezrin, radixin, and moesin) protein family. This affects cytoskeletal dynamics and the formation or stabilization of different membrane structures, such as filopodia, invadopodia, or ruffles.^(5–7)

PDPN is a specific marker for type I alveolar cells of the lung.^(2,8) Therefore, anti-PDPN monoclonal antibodies (mAbs) are useful in distinguishing type I alveolar cells of the lung from the type II cells. We characterized the PDPNs of various animal species using specific anti-PDPN mAbs. We established antimouse (PMAb-1),⁽⁹⁾ antirat (PMAb-2),⁽¹⁰⁾ antirabbit (PMAb-32),⁽¹¹⁾ antidog (PMAb-38)⁽¹²⁾ and PMAb-48⁽¹³⁾, antibovine (PMAb-44),⁽¹⁴⁾ and anticat (PMAb-52)⁽¹⁵⁾ PDPN antibodies. PMAb-38 was found to be cancer cell specific since it did not react with lymphatic endothelial cells

and type I alveolar cells,⁽¹²⁾ but reacted with squamous cell carcinomas⁽¹⁶⁾ and melanomas.⁽¹⁷⁾ We developed several antihuman PDPN mAbs, including NZ-1.2,⁽⁹⁾ LpMAb-7,^(18,19) LpMAb-10,⁽²⁰⁾ LpMAb-13,⁽²¹⁾ and LpMAb-17.⁽²²⁾ We also produced antiglycopeptide mAbs (GpMAbs) against human PDPN, namely, LpMAb-3,⁽¹⁸⁾ LpMAb-9,⁽¹⁸⁾ LpMAb-12,⁽²³⁾ LpMAb-19,⁽²⁴⁾ and LpMAb-21.^(25,26) Cancer-specific mAbs (CasMAbs) against human PDPN, such as LpMAb-2^(18,27) and LpMAb-23,^(28,29) were also established. Because PDPN is expressed in several normal tissues, CasMAbs against human PDPN should be utilized for clinical applications.

Antisheep PDPN mAbs have not been reported until date. In this study, we used immunohistochemical analyses to investigate the potential cross-reactions between anti-PDPN mAbs established for several species, particularly for sheep PDPN.

Materials and Methods

Immunohistochemical analyses

Normal sheep lungs were collected from two sheep after autopsy at Hokkaido University, fixed in 10% neutral-buffered formalin, and processed routinely to make paraffin-embedded tissue sections. Histological sections of thickness 4 μm were directly autoclaved in EnVision FLEX Target Retrieval Solution, high pH (Agilent Technologies, Inc.,

¹Department of Antibody Drug Development, Tohoku University Graduate School of Medicine, Sendai, Japan.

²New Industry Creation Hatchery Center, Tohoku University, Sendai, Japan.

³Laboratory of Comparative Pathology, Faculty of Veterinary Medicine, Hokkaido University, Sapporo, Japan.

⁴Laboratory of Infectious Diseases, Department of Disease Control, Faculty of Veterinary Medicine, Hokkaido University, Sapporo, Japan.

⁵Department of Advanced Pharmaceutics, Faculty of Veterinary Medicine, Hokkaido University, Sapporo, Japan.

Santa Clara, CA) for 20 minutes. After blocking with SuperBlock T20 (PBS) Blocking Buffer (Thermo Fisher Scientific, Inc. Waltham, MA), sections were incubated with PMab-44 (10 μ g/mL) for 1 hour at room temperature and treated using Envision+ Kit (Agilent Technologies, Inc.) for 30 minutes. Then color was developed using 3,3'-diaminobenzidine tetrahydrochloride (DAB; Agilent Technologies, Inc.) for 2 minutes, and counterstaining was performed with hematoxylin (FUJIFILM Wako Pure Chemical Corporation, Osaka, Japan).

Results and Discussion

Previously, we have developed a mouse antibovine PDPN mAb, PMab-44 (IgG₁, kappa), which specifically detects bovine PDPN in immunohistochemical analyses.⁽¹⁴⁾ Furthermore, a series of bovine PDPN deletion or point mutants were used for investigating the binding epitopes of PMab-44

using flow cytometry and Western blotting.⁽³⁰⁾ The PLAG3 of bovine PDPN was identified as the PMab-44 critical epitope. A comparison of amino acid sequences revealed an 85% homology between bovine and sheep PDPN (Supplementary Fig. S1). Therefore, we further investigated whether PMab-44 cross-reacts with sheep PDPN.

Flow cytometry indicated that PMab-44 reacted with CHO-K1 cells that overexpressed sheep PDPN (CHO/sPDPN) (Supplementary Fig. S2). Other anti-PDPN mAbs, such as antimouse (PMab-1),⁽⁹⁾ antirat (PMab-2),⁽¹⁰⁾ anti-rabbit (PMab-32),⁽¹¹⁾ antidog PMab-38⁽¹²⁾ and PMab-48⁽¹³⁾, and anticat (PMab-52⁽¹⁵⁾), did not react with CHO/sPDPN (data not shown). These results indicate that only PMab-44 is useful for the detection of sheep PDPN.

Furthermore, we investigated the expression of sheep PDPN in sheep lungs using immunohistochemical analyses. For this purpose, we used lungs obtained from two sheep (No. 1 and No. 2). Both lungs were stained using PMab-44

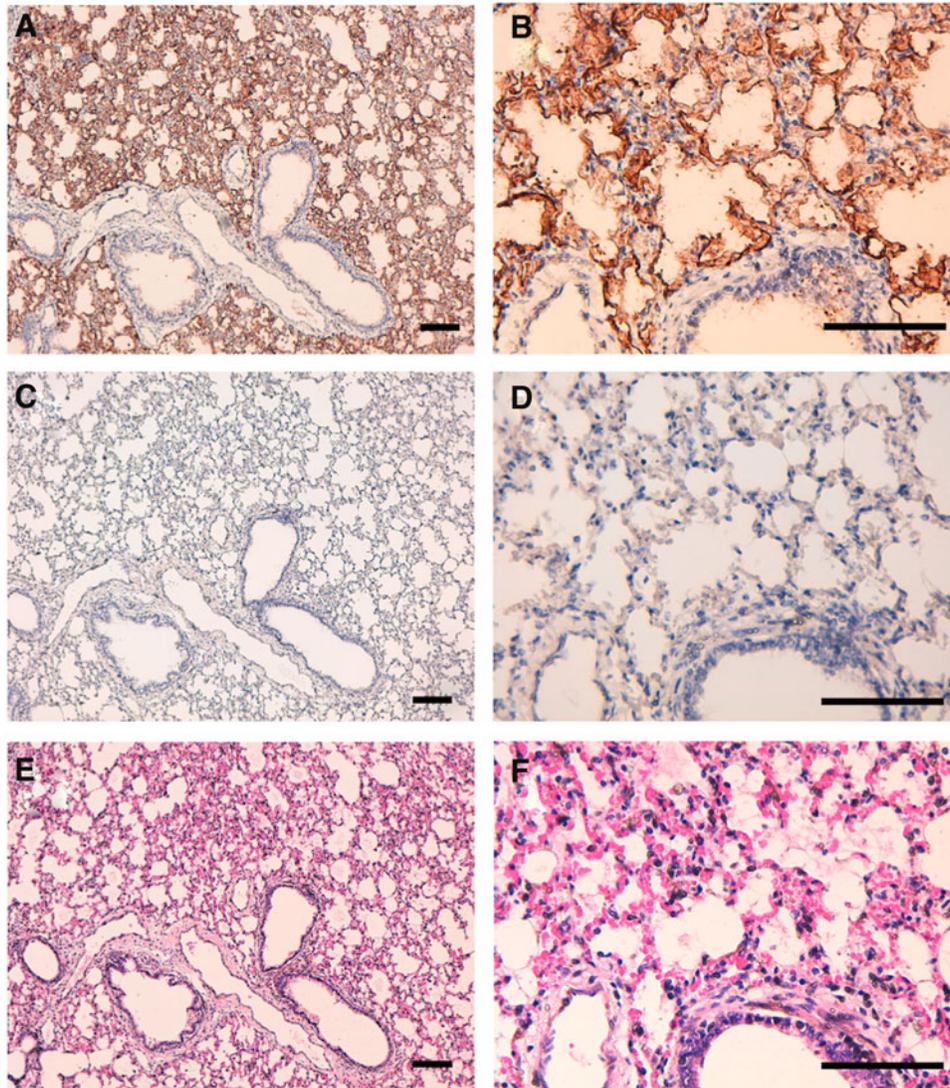


FIG. 1. Immunohistochemical analyses using sheep tissues. Histological sections of the sheep tissues were directly autoclaved in EnVision FLEX Target Retrieval Solution, high pH, for 20 minutes. After blocking, the sections were incubated with 10 μ g/mL of PMab-44 (A, B) or with blocking buffer (C, D), followed by detection using Envision+ Kit. (E, F) Hematoxylin and eosin staining. Scale bar = 100 μ m.

(10 µg/mL) when EnVision FLEX Target Retrieval Solution, high pH, was used for an antigen retrieval procedure. Representative results (No. 2) are depicted in Figure 1. Type I alveolar cells, but not the bronchial cells, were strongly stained with PMab-44 (Fig. 1A, B). Staining was not observed without PMab-44 (Fig. 1C, D). The identical tissue (No. 2) was stained moderately using PMab-44 (1 µg/mL) when citrate buffer (pH 6) was used for antigen retrieval (Supplementary Fig. S3), indicating that a specific staining by PMab-44 was observed at a low concentration of PMab-44 (1 µg/mL) or under universal antigen retrieval conditions (i.e., when citrate buffer was used). Unfortunately, the sheep kidney and colon were not stained by PMab-44 in this study (data not shown). This may be attributed to the varying expression levels of sheep PDPN among different tissues.

In conclusion, the antibovine PDPN mAb PMab-44 is useful for the detection of sheep PDPN in sheep lungs using immunohistochemical analyses. Further studies are necessary to demonstrate the capacity of PMab-44 for detection of sheep PDPN in other tissues.

Acknowledgments

This research was supported, in part, by AMED under grant numbers: JP18am0101078 (Y.K.), JP18am0301010 (Y.K.), and JP18ae0101028 (Y.K.), and by JSPS KAKENHI grant number 17K07299 (M.K.K.) and grant number 16K10748 (Y.K.).

Author Disclosure Statement

No competing financial interests exist.

References

- Kaneko MK, Kato Y, Kitano T, and Osawa M: Conservation of a platelet activating domain of Aggrus/podoplanin as a platelet aggregation-inducing factor. *Gene* 2006;378:52–57.
- Kato Y, Fujita N, Kunita A, Sato S, Kaneko M, Osawa M, and Tsuruo T: Molecular identification of Aggrus/T1alpha as a platelet aggregation-inducing factor expressed in colorectal tumors. *J Biol Chem* 2003;278:51599–51605.
- Kaneko M, Kato Y, Kunita A, Fujita N, Tsuruo T, and Osawa M: Functional sialylated O-glycan to platelet aggregation on Aggrus (T1alpha/podoplanin) molecules expressed in Chinese Hamster Ovary cells. *J Biol Chem* 2004; 279:38838–38843.
- Sekiguchi T, Takemoto A, Takagi S, Takatori K, Sato S, Takami M, and Fujita N: Targeting a novel domain in podoplanin for inhibiting platelet-mediated tumor metastasis. *Oncotarget* 2016;7:3934–3946.
- Martin-Villar E, Megias D, Castel S, Yurrita MM, Vilaro S, and Quintanilla M: Podoplanin binds ERM proteins to activate RhoA and promote epithelial-mesenchymal transition. *J Cell Sci* 2006;119:4541–4553.
- Martin-Villar E, Fernandez-Munoz B, Parsons M, Yurrita MM, Megias D, Perez-Gomez E, Jones GE, and Quintanilla M: Podoplanin associates with CD44 to promote directional cell migration. *Mol Biol Cell* 2010;21:4387–4399.
- Martin-Villar E, Borda-d'Agua B, Carrasco-Ramirez P, Renart J, Parsons M, Quintanilla M, and Jones GE: Podoplanin mediates ECM degradation by squamous carcinoma cells through control of invadopodia stability. *Oncogene* 2015;34:4531–4544.
- Breiteneder-Geleff S, Soleiman A, Kowalski H, Horvat R, Amann G, Kriehuber E, Diem K, Weninger W, Tschachler E, Alitalo K, and Kerjaschki D: Angiosarcomas express mixed endothelial phenotypes of blood and lymphatic capillaries: Podoplanin as a specific marker for lymphatic endothelium. *Am J Pathol* 1999;154:385–394.
- Kaji C, Tsujimoto Y, Kato Kaneko M, Kato Y, and Sawa Y: Immunohistochemical examination of novel rat monoclonal antibodies against mouse and human podoplanin. *Acta Histochem Cytochem* 2012;45:227–237.
- Oki H, Honma R, Ogasawara S, Fujii Y, Liu X, Takagi M, Kaneko MK, and Kato Y: Development of sensitive monoclonal antibody PMab-2 against rat podoplanin. *Monoclon Antib Immunodiagn Immunother* 2015;34:396–403.
- Honma R, Fujii Y, Ogasawara S, Oki H, Liu X, Nakamura T, Kaneko MK, Takagi M, and Kato Y: Establishment of a novel monoclonal antibody PMab-32 against rabbit podoplanin. *Monoclon Antib Immunodiagn Immunother* 2016; 35:41–47.
- Honma R, Kaneko MK, Ogasawara S, Fujii Y, Konnai S, Takagi M, and Kato Y: Specific detection of dog podoplanin expressed in renal glomerulus by a novel monoclonal antibody PMab-38 in immunohistochemistry. *Monoclon Antib Immunodiagn Immunother* 2016;35:212–216.
- Yamada S, Itai S, Kaneko MK, and Kato Y: PMab-48 recognizes dog podoplanin of lymphatic endothelial cells. *Monoclon Antib Immunodiagn Immunother* 2018;37: 63–66.
- Honma R, Ogasawara S, Kaneko M, Fujii Y, Oki H, Nakamura T, Takagi M, Konnai S, and Kato Y: PMab-44 detects bovine podoplanin in immunohistochemistry. *Monoclon Antib Immunodiagn Immunother* 2016;35:186–190.
- Yamada S, Itai S, Nakamura T, Yanaka M, Saidoh N, Chang YW, Handa S, Harada H, Kagawa Y, Ichii O, Konnai S, Kaneko MK, and Kato Y: PMab-52: Specific and sensitive monoclonal antibody against cat podoplanin for immunohistochemistry. *Monoclon Antib Immunodiagn Immunother* 2017;36:224–230.
- Kaneko MK, Honma R, Ogasawara S, Fujii Y, Nakamura T, Saidoh N, Takagi M, Kagawa Y, Konnai S, and Kato Y: PMab-38 recognizes canine podoplanin of squamous cell carcinomas. *Monoclon Antib Immunodiagn Immunother* 2016;35:263–266.
- Ogasawara S, Honma R, Kaneko MK, Fujii Y, Kagawa Y, Konnai S, and Kato Y: Podoplanin expression in canine melanoma. *Monoclon Antib Immunodiagn Immunother* 2016;35:304–306.
- Kato Y, and Kaneko MK: A cancer-specific monoclonal antibody recognizes the aberrantly glycosylated podoplanin. *Sci Rep* 2014;4:5924.
- Kato Y, Kunita A, Abe S, Ogasawara S, Fujii Y, Oki H, Fukayama M, Nishioka Y, and Kaneko MK: The chimeric antibody chLpMab-7 targeting human podoplanin suppresses pulmonary metastasis via ADCC and CDC rather than via its neutralizing activity. *Oncotarget* 2015;6:36003–36018.
- Ogasawara S, Oki H, Kaneko MK, Hozumi Y, Liu X, Honma R, Fujii Y, Nakamura T, Goto K, Takagi M, and Kato Y: Development of monoclonal antibody LpMab-10 recognizing non-glycosylated PLAG1/2 domain including Thr34 of human podoplanin. *Monoclon Antib Immunodiagn Immunother* 2015;34:318–326.

21. Ogasawara S, Kaneko MK, Honma R, Oki H, Fujii Y, Takagi M, Suzuki H, and Kato Y: Establishment of mouse monoclonal antibody LpMab-13 against human podoplanin. *Monoclon Antib Immunodiagn Immunother* 2016;35:155–162.
22. Kato Y, Ogasawara S, Oki H, Honma R, Takagi M, Fujii Y, Nakamura T, Saidoh N, Kanno H, Umetsu M, Kamata S, Kubo H, Yamada M, Sawa Y, Morita K, Harada H, Suzuki H, and Kaneko MK: Novel monoclonal antibody LpMab-17 developed by CasMab technology distinguishes human podoplanin from monkey podoplanin. *Monoclon Antib Immunodiagn Immunother* 2016;35:109–116.
23. Kato Y, Ogasawara S, Oki H, Goichberg P, Honma R, Fujii Y, and Kaneko MK: LpMab-12 established by CasMab technology specifically detects sialylated O-Glycan on Thr52 of platelet aggregation-stimulating domain of human podoplanin. *PLoS One* 2016;11:e0152912.
24. Ogasawara S, Kaneko MK, and Kato Y: LpMab-19 recognizes sialylated O-Glycan on Thr76 of human podoplanin. *Monoclon Antib Immunodiagn Immunother* 2016;35:245–253.
25. Kaneko MK, Nakamura T, Honma R, Ogasawara S, Fujii Y, Abe S, Takagi M, Harada H, Suzuki H, Nishioka Y, and Kato Y: Development and characterization of antiglycopeptide monoclonal antibodies against human podoplanin, using glycan-deficient cell lines generated by CRISPR/Cas9 and TALEN. *Cancer Med* 2017;6:382–396.
26. Kato Y, Kunita A, Fukayama M, Abe S, Nishioka Y, Uchida H, Tahara H, Yamada S, Yanaka M, Nakamura T, Saidoh N, Yoshida K, Fujii Y, Honma R, Takagi M, Ogasawara S, Murata T, and Kaneko MK: Antiglycopeptide mouse monoclonal antibody LpMab-21 exerts antitumor activity against human podoplanin through antibody-dependent cellular cytotoxicity and complement-dependent cytotoxicity. *Monoclon Antib Immunodiagn Immunother* 2017;36:20–24.
27. Kaneko MK, Yamada S, Nakamura T, Abe S, Nishioka Y, Kunita A, Fukayama M, Fujii Y, Ogasawara S, and Kato Y: Antitumor activity of chLpMab-2, a human–mouse chimeric cancer-specific antihuman podoplanin antibody, via antibody-dependent cellular cytotoxicity. *Cancer Med* 2017;6:768–777.
28. Yamada S, Kaneko MK, and Kato Y: LpMab-23: A cancer-specific monoclonal antibody against human podoplanin. *Monoclon Antib Immunodiagn Immunother* 2017;36:72–76.
29. Kaneko MK, Nakamura T, Kunita A, Fukayama M, Abe S, Nishioka Y, Yamada S, Yanaka M, Saidoh N, Yoshida K, Fujii Y, Ogasawara S, and Kato Y: ChLpMab-23: Cancer-specific human-mouse chimeric anti-podoplanin antibody exhibits antitumor activity via antibody-dependent cellular cytotoxicity. *Monoclon Antib Immunodiagn Immunother* 2017;36:104–112.
30. Yamada S, Honma R, Kaneko MK, Nakamura T, Yanaka M, Saidoh N, Takagi M, Konnai S, and Kato Y: Characterization of the anti-bovine podoplanin monoclonal antibody PMab-44. *Monoclon Antib Immunodiagn Immunother* 2017;36:129–134.

Address correspondence to:

Yukinari Kato
New Industry Creation Hatchery Center
Tohoku University
2-1, Seiryomachi, Aoba-ku
Sendai, Miyagi 980-8575
Japan

E-mail: yukinarikato@med.tohoku.ac.jp

Received: September 16, 2018

Accepted: November 25, 2018