

# LpMab-23: A Cancer-Specific Monoclonal Antibody Against Human Podoplanin

Shinji Yamada,<sup>1,2</sup> Satoshi Ogasawara,<sup>2</sup> Mika K. Kaneko,<sup>1,2</sup> and Yukinari Kato<sup>1-3</sup>

Human podoplanin (hPDPN), the ligand of C-type lectin-like receptor-2, is involved in cancer metastasis. Until now, many monoclonal antibodies (mAbs) have been established against hPDPN. However, it is still difficult to develop a cancer-specific mAb (CasMab) against hPDPN because the protein sequence of hPDPN expressed in cancer cells is the same as that in normal cells. Herein, we report LpMab-23 of the mouse IgG<sub>1</sub> subclass, a novel CasMab against hPDPN. In an immunohistochemical analysis, LpMab-23 reacted with tumor cells of human oral cancer, but did not react with normal cells such as lymphatic endothelial cells (LECs). In contrast, LpMab-17, another anti-hPDPN mAb, reacted with both tumor cells and LECs. Furthermore, flow cytometric analysis revealed that LpMab-23 reacted with hPDPN-expressing cancer cell lines (LN319, RERF-LC-AI/hPDPN, Y-MESO-14/hPDPN, and HSC3/hPDPN) but showed little reaction with normal cells (LECs and HEK-293T), although another anti-hPDPN mAb, LpMab-7, reacted with both hPDPN-expressing cancer cells and normal cells, indicating that LpMab-23 is a CasMab against hPDPN.

**Keywords:** podoplanin, monoclonal antibody, cancer-specific antibody, CasMab

## Introduction

**H**UMAN PODOPLANIN (hPDPN) is expressed in many cancers, including brain tumors, lung cancers, esophageal cancers, osteosarcomas, and malignant mesotheliomas.<sup>(1-13)</sup> C-type lectin-like receptor 2 is an endogenous receptor of hPDPN and is involved in platelet aggregation.<sup>(14-16)</sup> Recently, hPDPN was reported to be a potential biomarker for identifying the presence of early infiltrative esophageal squamous cell carcinoma.<sup>(17)</sup> Three different patterns can be distinguished in the expression of hPDPN in the basal layer of squamous epithelium lesions as follows: complete type, noncontinuous type, or missing type. The diagnosis of high-grade intraepithelial neoplasia can be made if the basal layer shows noncontinuous or complete expression of hPDPN. In contrast, the diagnosis of early infiltration can be made if hPDPN expression is completely missing. In these studies, D2-40, one of the anti-hPDPN monoclonal antibodies (mAbs), was used.

Although many anti-hPDPN mAbs have been reported, almost all the mAbs such as D2-40 and NZ-1 react with the peptides of hPDPN.<sup>(4,18-22)</sup> In contrast, we have used our original technology to produce anti-glycopeptide mAbs

(GpMabs) against hPDPN.<sup>(23)</sup> One of these GpMabs, LpMab-2, was shown to be a cancer-specific mAb (CasMab). LpMab-2 recognizes both an aberrant *O*-glycosylation and a Thr55-Leu64 peptide from hPDPN. Because LpMab-2 reacts with hPDPN-expressing cancer cells but not with normal cells, as shown by flow cytometry and immunohistochemistry, it is expected to be useful for molecular targeting therapy against hPDPN-expressing cancers. Nevertheless, LpMab-2 is not very useful in immunohistochemical analysis; therefore, another CasMab against hPDPN is necessary. In this study, we report LpMab-23, the second CasMab against hPDPN, which is very useful in immunohistochemistry.

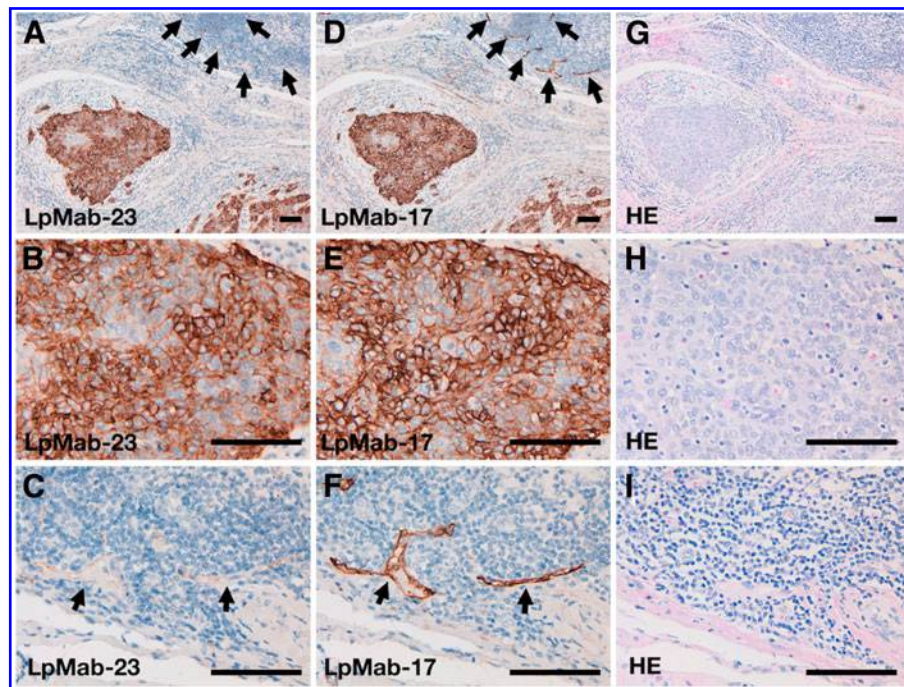
## Materials and Methods

### Hybridoma production

Female BALB/c mice (4 weeks old) were purchased from CLEA Japan (Tokyo, Japan). Animals were housed in a specific pathogen-free environment. The Animal Care and Use Committee of Tohoku University approved the animal experiments described herein. BALB/c mice were immunized by intraperitoneal (i.p.) injection of  $1 \times 10^8$  LN229/

Departments of <sup>1</sup>Antibody Drug Development, and <sup>2</sup>Regional Innovation, Tohoku University Graduate School of Medicine, Sendai, Miyagi, Japan.

<sup>3</sup>New Industry Creation Hatchery Center, Tohoku University, Sendai, Miyagi, Japan.



**FIG. 1.** Immunohistochemical analysis using anti-hPDPN mAbs to detect the expression of hPDPN in oral cancers. Tissue sections were prepared from the oral squamous cell carcinoma. Sections were incubated with LpMab-23 (A–C) and LpMab-17 (D–F) and made to react with the Envision+ kit. Color was developed using 3, 3-diaminobenzidine tetrahydrochloride, and the samples were then counterstained with hematoxylin. Sections were stained with hematoxylin and eosin as well (G–I). Cancer cells were detected by both LpMab-23 (A, B) and LpMab-17 (D, E). LpMab-23 showed little reaction with LECs (A, C). Arrows: LECs. Scale bar = 100  $\mu$ m. hPDPN, human podoplanin; LECs, lymphatic endothelial cells; mAbs, monoclonal antibodies.

hPDPN cells together with Imject Alum (Thermo Fisher Scientific, Inc., Waltham, MA). LN229/hPDPN cells were produced previously.<sup>(23)</sup> After several additional immunizations, a booster injection was given i.p. 2 days before spleen cells were harvested. The spleen cells were fused with P3U1 cells (ATCC, Manassas, VA) using PEG1500 (Roche Diagnostics, Indianapolis, IN). The hybridomas were grown in the RPMI medium (Wako Pure Chemical Industries, Ltd., Osaka, Japan) with hypoxanthine, aminopterin, and thymidine selection medium supplement (Thermo Fisher Scientific, Inc.). Using enzyme-linked immunosorbent assay (ELISA), the culture supernatants were screened for binding to recombinant hPDPN purified from LN229/hPDPN cells.

#### Immunohistochemical analyses

This study examined oral cancer patients who underwent surgery at the Sendai Medical Center as described previously.<sup>(24)</sup> Informed consent for obtaining samples and for subsequent data analyses was obtained from patients or the patient's guardian, and 4- $\mu$ m-thick histologic sections were deparaffinized in xylene and rehydrated. After antigen retrieval using the Envision FLEX system (high pH; Dako, Glostrup, Denmark), sections were incubated with 10  $\mu$ g/mL of LpMab-23 or 1  $\mu$ g/mL of LpMab-17 for 1 hour at room temperature followed by treatment with Envision+ kit (Dako) for 30 minutes. Color was developed using 3, 3-diaminobenzidine tetrahydrochloride (DAB; Dako) for 2 minutes; subsequently, the sections were counterstained with hematoxylin (Wako Pure Chemical Industries Ltd.).

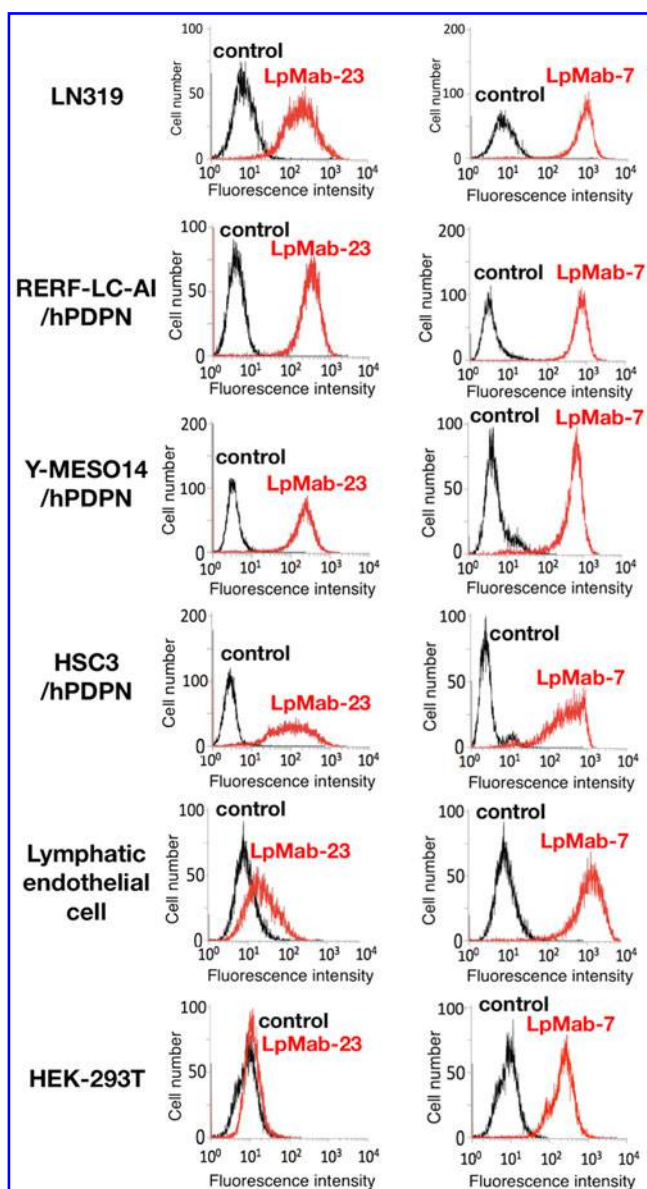
#### Flow cytometry

Cell lines in this study (LN319 of glioblastoma, RERF-LC-AI/hPDPN of lung cancer, Y-MESO-14/hPDPN of mesothelioma, HSC3/hPDPN of oral cancer, lymphatic endothelial cell (LEC), and HEK-293T of renal epithelial cell) were cultured as described previously.<sup>(23)</sup> Cell lines were harvested by brief exposure to 0.25% trypsin/1 mM EDTA (Nacalai Tesque, Inc., Kyoto, Japan). After washing, the cells were treated with LpMab-23 or LpMab-7 (1  $\mu$ g/mL) for 30 minutes at 4°C, followed by treatment with Oregon Green 488 goat anti-mouse IgG (Thermo Fisher Scientific, Inc.). Fluorescence data were collected using a Cell Analyzer EC800 (Sony Corp., Tokyo, Japan).

#### Results and Discussion

We immunized one mouse with the LN229/hPDPN, and the harvested spleen cells were fused with P3U1. ELISA screening was performed with supernatants from 960 hybridomas. Among 135 ELISA-positive wells, 19 wells reacted with LN229/hPDPN, but not with LN229 in flow cytometry (data not shown). We performed single-cell cloning for 19 wells by limiting dilution and could obtain 14 hybridomas. Until now, six clones have been reported, including LpMab-10, LpMab-12, LpMab-13, LpMab-17, LpMab-19, and LpMab-21.<sup>(23–33)</sup> In this study, we report the new clone, LpMab-23 (IgG<sub>1</sub>, kappa).

To investigate whether LpMab-23 is a CasMab, we performed immunohistochemical analyses using oral cancers.



**FIG. 2.** Flow cytometric analysis by anti-hPDPN mAbs. Cells were treated with LpMab-23, LpMab-7 (1  $\mu$ g/mL; red), or control phosphate-buffered saline (black) for 30 minutes at 4°C, followed by treatment with anti-mouse IgG-Oregon green. Fluorescence data were collected using a Cell Analyzer EC800.

As shown in Figure 1A and B, LpMab-23 reacted with tumor cells, which were also detected by another anti-hPDPN mAb, LpMab-17 (Fig. 1D, E).<sup>(24)</sup> In contrast, LpMab-23 showed little reaction with LECs (Fig. 1A, C), which were recognized by LpMab-17 (Fig. 1D, F). We further performed immunohistochemical analysis using 10 cases of oral cancers and obtained the same results (data not shown), indicating that LpMab-23 is a CasMab against hPDPN.

Flow cytometric analysis showed that LpMab-23 reacted with hPDPN-expressing cancer cell lines (LN319 of glioblastoma, RERF-LC-AI/hPDPN of lung cancer, Y-MESO-14/hPDPN of mesothelioma, and HSC3/hPDPN of oral cancer) but showed little reaction with normal cells (LECs

and HEK-293T of a renal epithelial cell) (Fig. 2). In contrast, an anti-hPDPN mAb, LpMab-7, reacted with both hPDPN-expressing cancer cells and normal cells, indicating that LpMab-23 is cancer specific.

Taken together, these results show that LpMab-23 might be useful for antibody therapy against hPDPN-expressing cancers. Reports have indicated that hPDPN is expressed in many normal cells<sup>(34,35)</sup>; therefore, anti-hPDPN mAbs might trigger unexpected side effects. To ensure the safety and efficacy of anti-hPDPN mAbs, CasMabs such as LpMab-23 should be applied clinically to cancer patients.

### Acknowledgments

We thank Takuro Nakamura, Miyuki Yanaka, Noriko Saidoh, and Kanae Yoshida for their excellent technical assistance. We also thank Yuki Fujii, Hiroaki Uchida, and Hideaki Tahara for their specialized advice. This work was supported, in part, by the Basic Science and Platform Technology Program for Innovative Biological Medicine from Japan Agency for Medical Research and development, AMED (Y.K.), by the Translational Research Network Program from AMED (Y.K.), by Project for utilizing glycans in the development of innovative drug discovery technologies from AMED (Y.K.), by the Platform for Drug Discovery, Informatics, and Structural Life Science (PDIS) from AMED (Y.K.), by JSPS KAKENHI Grant No. 26440019 (M.K.K.) and 16K10748 (Y.K.), and by the Regional Innovation Strategy Support Program from the Ministry of Education, Culture, Sports, Science and Technology (MEXT) of Japan (Y.K.). This work was performed, in part, under the Cooperative Research Program of Institute for Protein Research, Osaka University, CR-16-05 and by the Grant for Joint Research Project of the Institute of Medical Science, the University of Tokyo. We thank Enago ([www.enago.jp](http://www.enago.jp)) for the English language review.

### Author Disclosure Statement

No competing financial interests exist.

### References

- Martin-Villar E, Scholl FG, Gamallo C, Yurrita MM, Munoz-Guerra M, Cruces J, and Quintanilla M: Characterization of human PA2.26 antigen (T1alpha-2, podoplanin), a small membrane mucin induced in oral squamous cell carcinomas. *Int J Cancer* 2005;113:899–910.
- Yuan P, Temam S, El-Naggar A, Zhou X, Liu D, Lee J, and Mao L: Overexpression of podoplanin in oral cancer and its association with poor clinical outcome. *Cancer* 2006;107:563–569.
- Kunita A, Kashima TG, Ohazama A, Grigoriadis AE, and Fukayama M: Podoplanin is regulated by AP-1 and promotes platelet aggregation and cell migration in osteosarcoma. *Am J Pathol* 2011;179:1041–1049.
- Kato Y, Kaneko MK, Kuno A, Uchiyama N, Amano K, Chiba Y, Hasegawa Y, Hirabayashi J, Narimatsu H, Mishima K, and Osawa M: Inhibition of tumor cell-induced platelet aggregation using a novel anti-podoplanin antibody reacting with its platelet-aggregation-stimulating domain. *Biochem Biophys Res Commun* 2006;349:1301–1307.
- Mishima K, Kato Y, Kaneko MK, Nakazawa Y, Kunita A, Fujita N, Tsuruo T, Nishikawa R, Hirose T, and Matsutani

- M: Podoplanin expression in primary central nervous system germ cell tumors: A useful histological marker for the diagnosis of germinoma. *Acta Neuropathol* 2006;111:563–568.
6. Mishima K, Kato Y, Kaneko MK, Nishikawa R, Hirose T, and Matsutani M: Increased expression of podoplanin in malignant astrocytic tumors as a novel molecular marker of malignant progression. *Acta Neuropathol* 2006;111:483–488.
  7. Abe S, Morita Y, Kaneko MK, Hanibuchi M, Tsujimoto Y, Goto H, Kakiuchi S, Aono Y, Huang J, Sato S, Kishuku M, Taniguchi Y, Azuma M, Kawazoe K, Sekido Y, Yano S, Akiyama S, Sone S, Minakuchi K, Kato Y, and Nishioka Y: A novel targeting therapy of malignant mesothelioma using anti-podoplanin antibody. *J Immunol* 2013;190:6239–6249.
  8. Kato Y, Vaidyanathan G, Kaneko MK, Mishima K, Srivastava N, Chandramohan V, Pegram C, Keir ST, Kuan CT, Bigner DD, and Zalutsky MR: Evaluation of anti-podoplanin rat monoclonal antibody NZ-1 for targeting malignant gliomas. *Nucl Med Biol* 2010;37:785–794.
  9. Kato Y, Fujita N, Kunita A, Sato S, Kaneko M, Osawa M, and Tsuruo T: Molecular identification of Aggrus/T1alpha as a platelet aggregation-inducing factor expressed in colorectal tumors. *J Biol Chem* 2003;278:51599–51605.
  10. Kato Y, Sasagawa I, Kaneko M, Osawa M, Fujita N, and Tsuruo T: Aggrus: A diagnostic marker that distinguishes seminoma from embryonal carcinoma in testicular germ cell tumors. *Oncogene* 2004;23:8552–8556.
  11. Kato Y, Kaneko M, Sata M, Fujita N, Tsuruo T, and Osawa M: Enhanced expression of Aggrus (T1alpha/podoplanin), a platelet-aggregation-inducing factor in lung squamous cell carcinoma. *Tumour Biol* 2005;26:195–200.
  12. Kunita A, Kashima TG, Morishita Y, Fukayama M, Kato Y, Tsuruo T, and Fujita N: The platelet aggregation-inducing factor aggrus/podoplanin promotes pulmonary metastasis. *Am J Pathol* 2007;170:1337–1347.
  13. Kaneko MK, Kato Y, Kitano T, and Osawa M: Conservation of a platelet activating domain of Aggrus/podoplanin as a platelet aggregation-inducing factor. *Gene* 2006;378:52–57.
  14. Suzuki-Inoue K, Kato Y, Inoue O, Kaneko MK, Mishima K, Yatomi Y, Yamazaki Y, Narimatsu H, and Ozaki Y: Involvement of the snake toxin receptor CLEC-2, in podoplanin-mediated platelet activation, by cancer cells. *J Biol Chem* 2007;282:25993–26001.
  15. Kato Y, Kaneko MK, Kunita A, Ito H, Kameyama A, Ogasawara S, Matsuura N, Hasegawa Y, Suzuki-Inoue K, Inoue O, Ozaki Y, and Narimatsu H: Molecular analysis of the pathophysiological binding of the platelet aggregation-inducing factor podoplanin to the C-type lectin-like receptor CLEC-2. *Cancer Sci* 2008;99:54–61.
  16. Nagae M, Morita-Matsumoto K, Kato M, Kaneko MK, Kato Y, and Yamaguchi Y: A platform of C-Type lectin-like receptor CLEC-2 for binding O-glycosylated podoplanin and nonglycosylated rhodocytin. *Structure* 2014;22:1711–1721.
  17. Chen G, Xu R, Yue B, Mei X, Li P, Zhou X, Huang S, Gong L, and Zhang S: The expression of podoplanin protein is a diagnostic marker to distinguish the early infiltration of esophageal squamous cell carcinoma. *Oncotarget* 2017;8:19013–19020.
  18. Ogasawara S, Kaneko MK, Price JE, and Kato Y: Characterization of anti-podoplanin monoclonal antibodies: Critical epitopes for neutralizing the interaction between podoplanin and CLEC-2. *Hybridoma (Larchmt)* 2008;27:259–267.
  19. Takagi S, Sato S, Oh-hara T, Takami M, Koike S, Mishima Y, Hatake K, and Fujita N: Platelets promote tumor growth and metastasis via direct interaction between Aggrus/podoplanin and CLEC-2. *PLoS One* 2013;8:e73609.
  20. Nakazawa Y, Takagi S, Sato S, Oh-hara T, Koike S, Takami M, Arai H, and Fujita N: Prevention of hematogenous metastasis by neutralizing mice and its chimeric anti-Aggrus/podoplanin antibodies. *Cancer Sci* 2011;102:2051–2057.
  21. Marks A, Sutherland DR, Bailey D, Iglesias J, Law J, Lei M, Yeger H, Banerjee D, and Baumal R: Characterization and distribution of an oncofetal antigen (M2A antigen) expressed on testicular germ cell tumours. *Br J Cancer* 1999;80:569–578.
  22. Kono T, Shimoda M, Takahashi M, Matsumoto K, Yoshimoto T, Mizutani M, Tabata C, Okoshi K, Wada H, and Kubo H: Immunohistochemical detection of the lymphatic marker podoplanin in diverse types of human cancer cells using a novel antibody. *Int J Oncol* 2007;31:501–508.
  23. Kato Y, and Kaneko MK: A cancer-specific monoclonal antibody recognizes the aberrantly glycosylated podoplanin. *Sci Rep* 2014;4:5924.
  24. Kato Y, Ogasawara S, Oki H, Honma R, Takagi M, Fujii Y, Nakamura T, Saidoh N, Kanno H, Umetsu M, Kamata S, Kubo H, Yamada M, Sawa Y, Morita K, Harada H, Suzuki H, and Kaneko MK: Novel monoclonal antibody LpMab-17 developed by CasMab technology distinguishes human podoplanin from monkey podoplanin. *Monoclon Antib Immunodiagn Immunother* 2016;35:109–116.
  25. Oki H, Ogasawara S, Kaneko MK, Takagi M, Yamauchi M, and Kato Y: Characterization of monoclonal antibody LpMab-3 recognizing sialylated glycopeptide of podoplanin. *Monoclon Antib Immunodiagn Immunother* 2015;34:44–50.
  26. Oki H, Kaneko MK, Ogasawara S, Tsujimoto Y, Liu X, Sugawara M, Takakubo Y, Takagi M, and Kato Y: Characterization of a monoclonal antibody LpMab-7 recognizing non-PLAG domain of podoplanin. *Monoclon Antib Immunodiagn Immunother* 2015;34:174–180.
  27. Kaneko MK, Oki H, Hozumi Y, Liu X, Ogasawara S, Takagi M, Goto K, and Kato Y: Monoclonal antibody LpMab-9 recognizes O-glycosylated N-terminus of human podoplanin. *Monoclon Antib Immunodiagn Immunother* 2015;34:310–317.
  28. Ogasawara S, Oki H, Kaneko MK, Hozumi Y, Liu X, Honma R, Fujii Y, Nakamura T, Goto K, Takagi M, and Kato Y: Development of monoclonal antibody LpMab-10 recognizing non-glycosylated PLAG1/2 domain including Thr34 of human podoplanin. *Monoclon Antib Immunodiagn Immunother* 2015;34:318–326.
  29. Kaneko MK, Oki H, Ogasawara S, Takagi M, and Kato Y: Anti-podoplanin monoclonal antibody LpMab-7 detects metastatic regions of osteosarcoma. *Monoclon Antib Immunodiagn Immunother* 2015;34:154–161.
  30. Kato Y, Kunita A, Abe S, Ogasawara S, Fujii Y, Oki H, Fukayama M, Nishioka Y, and Kaneko MK: The chimeric antibody chLpMab-7 targeting human podoplanin suppresses pulmonary metastasis via ADCC and CDC rather than via its neutralizing activity. *Oncotarget* 2015;6:36003–36018.
  31. Ogasawara S, Kaneko MK, Honma R, Oki H, Fujii Y, Takagi M, Suzuki H, and Kato Y: Establishment of mouse monoclonal antibody LpMab-13 against human podoplanin. *Monoclon Antib Immunodiagn Immunother* 2016;35:155–162.



32. Ogasawara S, Kaneko MK, and Kato Y: LpMab-19 recognizes sialylated O-glycan on Thr76 of human podoplanin. *Monoclon Antib Immunodiagn Immunother* 2016;35: 245–253.
33. Kato Y, Kunita A, Fukayama M, Abe S, Nishioka Y, Uchida H, Tahara H, Yamada S, Yanaka M, Nakamura T, Saidoh N, Yoshida K, Fujii Y, Honma R, Takagi M, Ogasawara S, Murata T, and Kaneko MK: Antiglycopeptide mouse monoclonal antibody LpMab-21 exerts antitumor activity against human podoplanin through antibody-dependent cellular cytotoxicity and complement-dependent cytotoxicity. *Monoclon Antib Immunodiagn Immunother* 2017;36:20–24.
34. Breiteneder-Geleff S, Matsui K, Soleiman A, Meraner P, Poczewski H, Kalt R, Schaffner G, and Kerjaschki D: Podoplanin, novel 43-kd membrane protein of glomerular epithelial cells, is down-regulated in puromycin nephrosis. *Am J Pathol* 1997;151:1141–1152.
35. Breiteneder-Geleff S, Soleiman A, Kowalski H, Horvat R, Amann G, Kriehuber E, Diem K, Weninger W, Tschachler E, Alitalo K, and Kerjaschki D: Angiosarcomas express mixed endothelial phenotypes of blood and lymphatic capillaries: Podoplanin as a specific marker for lymphatic endothelium. *Am J Pathol* 1999;154:385–394.

Address correspondence to:  
*Dr. Yukinari Kato*  
*New Industry Creation Hatchery Center*  
*Tohoku University*  
*2-1 Seiryomachi, Aoba-ku*  
*Sendai*  
*Miyagi 980-8575*  
*Japan*

*E-mail: yukinari-k@bea.hi-ho.ne.jp*

*Received: January 17, 2017*

*Accepted: February 28, 2017*